SYNTHESIS OF ANTHOCYANIDINS—I

THE OXIDATIVE GENERATION OF FLAVYLIUM CATIONS USING BENZOQUINONES

J. G. SWEENY and G. A. IACOBUCCI*

Corporate Research and Development Department, The Coca-Cola Company, P.O. Drawer 1734, Atlanta, GA 30301, U.S.A.

(Received in USA 14 March 1977; Received in UK for publication 27 June 1977)

[Dedicated to Prof. ROBERT B. WOODWARD on the occasion of hir 60th birthday]

Abstract—The addition of benzoquinone and especially chloranil has been found to greatly enhance the yields of 3-deoxyanthocyanidins produced on oxidation of flavan-4-ols. In contrast, only slight improvements are noted when the same oxidants are used in the oxidation of the related flavan-3,4-diols.

Since the early contributions of Willstätter to the total synthesis of anthocyanidins and the more definitive work of Robinson that followed, several alternative approaches have been considered to facilitate the synthesis of the highly oxygenated, phenolic natural pigments.

Convenient transformations of related compounds like flavonols, flavanones and dihydroflavonols to yield flavylium salts have been sought, sometimes motivated by chemurgic interests, in other cases in search of plausible sequences of biosynthetic significance for this group of natural products.

Of particular synthetic interest is the report by Jurd² that the flav-2-ene 2 is readily oxidized to pentamethyl-cyanidin 3 in high yield, on heating with benzoquinone in acetic acid at 100°, thus facilitating the overall conversion of the flavonol 1 to the flavylium salt 3.

As a part of our own studies, we have explored in further detail the scope of this oxidation, by extending it

to a series of flavan-4-ols and flavan-3,4-diols of varied structure.

The flavan-4-ols shown in Table 1 were prepared by NaBH₄ reduction of the corresponding flavanones and yielded exclusively the 2.4-cis-conformers having the C₄-OH equatorial. As expected, inversion of C₄ occurred during the synthesis of 6 from 4 using EtOH/HOAc. The results shown in Table 1 confirm the effect of quinone addition upon anthocyanidin yield² and are best rationalized by assuming the intermediate formation of flav-3-enes, which make possible the subsequent abstraction of the C₂ allylic hydrogen by the quinone.

Compounds 7 and 10 failed to react under these conditions, suggesting that oxygen substitution at C_7 facilitates the formation of the intermediate flavene.

From a preparative point of view, the oxidation of a flavan-4-ol is preferred over that of a flavene, mainly because the former is directly available from the flavanone in one step, while the preparation of the latter

Scheme 1.

Table 1. Effect of benzoquinone and chloranil addition on the oxidation of flavan-4-ols to 3-deoxyanthocyanidins

		3-Deoxyanthocyanidin Yield (%) =		
	Compound	None	Benzoquinone	Chloranil
4	R ₁ = R ₂ = R ₅ = R ₆ = OCH ₃ ; R ₃ = H; R ₄ = OH	3	25	66
5	$R_1 = R_2 = R_6 = OCH_3$; $R_3 = R_5 = H$; $R_4 = OH$	0	31	59
6	R ₁ = R ₂ = R ₅ = R ₆ = OCH ₃ ; R ₃ = OC ₂ H ₅ ; R ₄ = H	6	22	69
7	R ₁ = R ₂ = R ₃ = R ₅ = R ₆ = H; R ₄ = OH	0	0	0
8	$R_2 = R_3 = R_5 = R_6 = H; R_1 = R_4 = OH$	1	25	49
10	R ₁ = R ₂ = R ₃ = R ₅ = H; R ₄ = R ₆ = OH OCH ₃	0	0	0
9	H ₃ CO → OCH ₃ OCH ₃	2	45	63
	·· J			

Reaction mixture consisted of 10 mg flavanol, 10 mg oxidant and 1 ml acetic acid, heated at 100°C for 30 minutes. Yields of 3 deoxyanthocyanidin estimated spectrophotometrically after chromatography on acid-treated polyclar, with reference to the appropriate pure standards in 0.01 N HCl: tetramethylluteolinidin chloride (≈ 476 nm = 38,000) for compounds 4, 6, and 9; trimethylapigeninidin chloride (≈ 460 nm = 38,000) for compound 5, 7-hydroxyflavylium chloride (≈ 440 nm = 32,700) for compound 8.

requires first a flavanone-to-chalcone conversion, followed by borohydride reduction and acid catalyzed cyclization.³ The flavene route is, of course, the method of choice for flavanones lacking oxygen functionality at C_7 .

The flavan-3,4-diols and their methyl ethers listed in Table 2 were prepared by borohydride reduction of tetramethyldihydroquercetin, separation of the isomeric

Table 2. Effect of benzoquinone and chloranil addition on the oxidation of flavan-3,4-diols and their ethers to anthocyanidins

		Anthocyanidin Yield (%) 8			
	Compound	None	Benzoquinone	Chlorani	
11	R ₁ = R ₃ = OH; R ₂ = H	13	17	21	
12	R ₁ = R ₃ = OAc; R ₂ = H	12	20	14	
13	R ₁ = H, R ₂ = OH; R ₃ = OCH ₃	9	24	27	
14	R ₁ = OH; R ₂ = H; R ₃ = OCH ₃	12	25	27	
15	R ₁ = OCH ₃ ; R ₂ = H; R ₃ = OCH ₃	6	24	28	

^{3.} Reaction mixture 10 mg flavanol, 10 mg oxidant, 0.5 ml acetic acid and 0.5 ml 6.N HCl², heated at 100º for 30 minutes. Anthocyanidin yields were estimated spectrophotometrically in 0.01 N HCl in MeOH, as indicated in Table I, using ⁶ 540 nm = 34,700².

C₄ alcohols, and methylation with MeI/Ag₂O (Experimental). In this series, the oxidations did not proceed unless HCl was added to the reaction mixture. Contrary to a previous observation,2 the addition of benzoquinone did not improve the yield of anthocyanidin formed from the diol 11 (Table 2). Moreover, the situation did not improve when chloranil was used instead. When the stronger oxidant DDQ was reacted with 11 in acetic acid at room temperature, the parent 3-hydroxyflavanone was regenerated, but no formation of flavylium cation was noted. Table 2 shows that the anthocyanidin yields are neither dependent on the stereochemistry of the oxygen at C_4 nor are affected by the nature of its substitution. Also, the nature of the oxygen function at C_3 seems unimportant. This is especially surprising in the case of the 3-methoxyderivatives 13, 14 and 15, which yield the more stable pentamethylcyanidin 3. In parallel experiments, this anthocyanidin was recovered unchanged (95% yield) after heating at 100°C for 30 min in acetic acid, acetic acid-6N HCl (5:1), and acetic acid-6N HCl (5:1) containing 2 equivalents of chloranil. As shown in Table 3, the stability of 3 in aqueous acid is two orders of magnitude higher than that of its tetramethyl analogue

Comparison of Tables 1 and 2 indicates that the addition to the flavan-4-ols of an oxygen function at C₃ results in lower yields of anthocyanidins upon quinone oxidation, with no advantage of chloranil over benzoquinone.

The known tendency of flavan-3,4-diols to condense with themselves or other flavans in the presence of strong acids,⁴ may be a factor in determining the differences in reactivity noted above.

Table 3. Stability of anthocyanidins in 0.01 N HCl solution

$$R_1$$
 R_2
 R_4
 R_5
 R_5

	Anthocyanidin	Half Life (days)	
16	R ₁ = R ₂ = R ₄ * R ₅ = OCH ₃ ; R ₃ = OH	0 04	
17	R ₁ = R ₂ = R ₃ = R ₄ = R ₅ = OH	0.5	
3	R ₁ = R ₂ = R ₃ = R ₄ = R ₅ = OCH ₃	6	
18	R ₁ = R ₂ = R ₄ = R ₅ = OCH ₃ , R ₃ = H	> 100	

Measured spectrophotometrically at 540 nm (3, 16 and 17) and 476 nm (18).

EXPERIMENTAL

M.ps are uncorrected. 'H NMR spectra were recorded in CDCl₃ unless otherwise stated. Chemical shifts are given in ppm downfield from TMS. Coupling constants (J) are in c/s. Abbreviations: s, singlet; b.s., broad singlet; d, doublet; t, triplet; m, multiplet. Chloranil (Eastman) was recrystallized from toluene before use. All other reagents were used as received from the supplier and were reagent grade. Microanalyses were performed by Galbraith Analytical Laboratories, Knoxville, Tenn.

Quinone oxidation of flavan-4-ols [Table 1]. The mixture consisted of 10 mg flavan, 10 mg oxidant and 1 ml HOAc. After heating for 30 min at 100° in air, the soln was cooled and diluted with 0.01 N HCl in MeOH to a volume of 10 ml. A one ml aliquot was then chromatographed on acid-treated polyclar 5 (15×3 cm column) using 0.01 N HCl as eluant. The anthocyanin band was collected, diluted to 100 ml with 0.01 N HCl, and its absorption measured at 476 nm ($\epsilon = 38,000$) for tetramethylluteolinidin chloride and 460 nm ($\epsilon = 38,000$) for trimethylapigeninidin chloride. 7-OH flavylium chloride was eluted from the column with 0.01 N HCl in MeOH-H₂O (1:1) and had $\lambda_{max}440$ nm ($\epsilon = 32,700$). It showed the reported shift to λ_{max} 370 nm at higher pH due to hydrolysis to the chalcone.

Quinone oxidation of flavan-3,4-diols [Table 2]. The mixture consisted of 10 mg flavanol, 10 mg oxidant, 0.5 ml HOAc and 0.1 ml 6 N HCl.² After heating at 100° for 30 min, the soln was diluted to 10 ml with 0.01 N HCl in MeOH. A one ml aliquot of this soln was then chromatographed on acid-treated polyclar⁵ (15 × 3 cm column) using 0.01 N HCl in MeOH as eluant. The red anthocyanin band was diluted to 100 ml with 0.01 N HCl in MeOH and the absorbance read at 540 m μ . The pentamethyl-cyanidin chloride (3) used as standard was synthesized via quinone oxidation of 2.²⁻⁷. Two recrystallizations from 6 N HCl gave m.p. 145-8° and $\lambda_{max} = 512$ nm (0.01 N HCl). (lit. 152°, $\lambda_{max} = 512$).

3',4',5,7 - Tetramethoxy - 2,3 - trans - 3,4 - cis - flavan - 3,4 - diol (11) was prepared via the peracid oxidation of 9 using one equivalent of m-chloroperbenzoic acid in methylene chloride, instead of peracetic acid as described by Clark-Lewis.³ The hydrolysis was then conducted as reported to give 11 (36%) m.p. 166-7° (lit.³ 165-7°). Acetylation (18 hr, Ac₂O/py) afforded the 2,3-trans-3,4-cis-diacetate (12) m.p. 159-60° (lit.³ 155-7°).

Methylation of 3',4',5,7-tetramethyldihydroquercetin. 3',4',5,7-Tetramethyldihydroquercetin⁹ (900 mg). Ag₂O (750 mg), MeI (0.33 ml) and DMF (6 ml) were shaken vigorously in a 25 ml stoppered-flask in the dark for 18 hrs, as described. ¹⁰ 6 ml CHCl₃ was then added and the soln filtered through celite. After wash-

ing the celite with an additional 10 ml CHCl3, the combined filtrates were washed with 2×20 ml H₂O, dried (Na₂SO₄) and evaporated. TLC (silica gel) using EtOAc-hexane (1:1) as moving phase showed two spots, which were separated by column chromatography on SilicAR CC-7. The first compound to elute was obtained as light yellow crystals from MeOH, 128 mg, m.p. 138-9°. It was identified as 2',3,4,4',6',α-hexamethoxychalcone (lit. 9 132-3°), by analysis of the MS (m/e 388), NMR (6-Me groups), UV ($\lambda_{max\,343}^{EiOH}$, ϵ = 22,400) and particularly the IR, which was identical to the published one.9 The second material to elute was identified as 2,3 - trans - 3,3',4',5,7 methyldihydroquercetin, 331 mg, m.p. 151-2° (MeOH). Clark-Lewis and Korytnyk¹¹ report m.p. 169-70° for the completely racemic material prepared by a different route. Although our compound was only slightly optically active $[\alpha_D] < 1^\circ$, the m.p. difference may be due to this factor. IR (KBr)µ: 5.92, 6.20, 6.34, 6.58, 6.80, 7.00, 7.89, 8.02, 8.21, 8.63, 9.00, 9.72, NMR: 3 H m at 6.93 (ArH), 2 H b.s. at 6.06 (ArH), 1 H d at 5.17 (J = 9, C_2H), 6 H s at 3.88 (OCH₃), 3 H s at 3.78 (OCH₃), 3 H s at 3.44 (OCH₃) and 1 H m at 3.95 (C₃H obscured by the OCH₃ absorptions). (Calc. for C₂₀H₂₂O₇: C, 64.17; H, 5.88. Found: C, 64.44; H, 6.13%).

Sodium borohydride reduction of 2,3 - trans - 3,3',4',5,7 pentamethyldihydroquercetin. While stirring at room temp, NaBH₄ was added to a soln of 400 mg pentamethyldihydroquercetin in 50 ml of THF/EtOH (1:1). After 30 min an additional 100 mg was added and stirring continued for a total of 1 hr. The soln was then poured into 150 ml of cold 0.5% HOAc and extracted with 2×50 ml CHCl₃. Drying (Na2SO4) and evaporating the CHCl3 layers gave an oil which showed two spots on TLC analysis [silica gel, EtOAc/hexane (1:1) elution]. The mixture was separated by column chromatography on silicAR CC-7 to give: 2, 3 - trans - 3,4 - trans -3,3',4',5.7 - pentamethoxyflavan - 4 - ol (13), 93 mg (23%), m.p. 157-8° (MeOH) (lit.3 155°), MS, IR, and NMR in complete agreement with those reported,3 and 2,3 - trans - 3,4 - cis -3,3',4',5,7 - pentamethoxyflavan - 4 - ol (14), 280 mg (70%), m.p. 119-20° (EtOAc-hexane). IR (KBr)µ: 2.87, 3.40, 6.18, 6.28, 6.59, 6.81, 7.89, 8.32, 8.70, 9.10, 9.71. MS m/e (rel. intensity): no molecular ion, 358 (87), 343 (37), 327 (100), 221 (85), 194 (67), 179 (43) and 151 (39). NMR: 3 H m at 6.90 (ArH), 2 H b.s. at 6.01 $(C_6 + C_8H)$, 1 H d at 5.05 (J = 4, C₄H), 1 H d at 4.95 (J = 10, C₂H), 6 H s at 3.84 (OCH₃), 3 H s at 3.78 (OCH₃), 3 H s at 3.66 (OCH₃), 1 H q at 3.44 (J = 4 and 10, C_3H) and 3 H s at 3.24 (OCH₃). (Calc. for C₂₀H₂₄O₇: C, 63.83; H, 6.38. Found: C, 64.02; H, 6.55%).

2,3 - trans - 3,4 - cis - 3,4,3',4',5,7 - Hexamethoxyflavan (15). 3',4',5,7 - Tetramethoxyflavan - 3,4 - diol (m.p. 172-8°, mixture of 3,4-cis and 3,4-trans isomers from NaBH₄ reduction of tetramethyldihydroquercetin¹²), 450 mg, was methylated with MeI (0.17 ml) and Ag₂O (375 mg) in DMF (3 ml) as described above for tetramethyldihydroquercetin. The product mixture showed three spots on TLC which were separated by column chromatography on silicAR CC-7 (20% EtOAc-hexane elution). The first material to elute was 2,3 - trans - 3,4 - cis - 3,4,3',4',5,7 hexamethoxyflavan (15) 145 mg (30%), m.p. 120-3° (MeOH) raised to 126-7° after 2 recrystallizations. IR (KBr)µ: 6.19, 6.28, 6.58, 6.82, 7.01, 7.88, 8.31, 8.70 and 9.08; MS m/e (rel. intensity): 388 (6.5), 358 (100), 327 (83), 194 (79), 57 (94), 55 (60) and 43 (93). NMR: 3 H m at 6.90 (ArH), 2 H b.s. at 6.02 (C₆ and C₈H), 1 H d at $5.12 (J = 10, C_2H)$, 1 H d at 4.78 $(J = 4, C_4H)$, 6 H s at 3.84 (OCH₂), 3 H s at 3.79 (OCH₃), 3 H s at 3.70 (OCH₃), 3 H s at 3.48 (OCH₃), 1 H m at \sim 3.40 (C₃H) and 3 H s at 3.14 (OCH₃). (Calc. for C₂₁H₂₆O₇: C, 64.61; H, 6.67. Found: C, 64.69; H, 6.72%) In addition, there was obtained 179 mg (38%) of 13 and 115 mg (25%) of 14.

2,4 - cis - 4 - Hydroxy - 3',4',5,7 - tetramethoxyflavan (4), m.p. 112.5-113.5° (lit. 13 114°); 2,4 - cis - 4 - hydroxy - 4',5,7 - trimethoxyflavan (5), m.p. 156-8° (lit. 13 159°); 2,4 - cis - 4 - hydroxyflavan (7), m.p. 147-8° (lit. 148-9.5°); and 2,4 - cis - 4,4' - dihydroxyflavan (10), m.p. 182-4° (lit. 13 18)° were prepared by NaBH₄ reduction of the corresponding flavanones in EtOH. The stereochemistry is based on the assignments of the original authors and comes from analysis of the NMR spectra. 16

2,4 - trans - 4 - Ethoxy - 3',4',5,7 - tetramethoxyflavan (6), oil, was prepared from 4 by refluxing with HOAc in EtOH.³

2,4 - cis - 4.7 - Dihydroxyflavan (8). 7-hydroxyflavanone¹⁵

(200 mg) was reduced with NaBH₄ (100 mg) in EtOH at room temp. The BH₄⁻ was added over 1 hr, then the mixture allowed to stir at room temp. for 18 hr. Pouring the mixture into H₂O, neutralizing with HOAc, extracting with CHCl₃ and evaporation of the CHCl₃ gave 2,4-cis-4,7-dihydroxyflavan as an amorphous solid. After recrystallization from EtOAc-hexane, there was obtained 61 mg (30%), m.p. 138-9° (lit. 15 42-6°). IR (KBr) μ : 2.90, 6.12, 6.78, 8.66, 8.91, 9.42, 9.71. NMR: 1 H b.s. at 8.2 (OH), 6 H b.s. at 7.2-7.5 (ArH), 1 H m at 6.5 (C₆H), 1 H m at 6.4 (C₈H), 2 H m at 5.0 (C₂H and C₄H) and 2 H m at 2.2-2.4 (C₃H). MS m/e (rel. intensity): 242 (4), 224 (100), 223 (97), 165 (10), 152 (7.4), 147 (39) and 115 (4.7). (Calc. for C₁₅H₁₄O₃: C, 74.38; H, 5.78. Found: C, 74.25; H, 5.60%).

4',5,7-Trimethoxyflavylium chloride (trimethylapigeninidin chloride). A mixture of 200 mg 5, 160 mg chloranil and 5 ml HOAc was heated at 100° with stirring for 30 min. After cooling in ice, the soln was diluted with 5 ml of 0.01 N HCl in MeOH and added to the top of an 18 × 2.5 cm polyclar column (acid treated).5 The anthocyanin band was eluted with 0.01 N HCl and the eluant freeze-dried. After washing with 2×10 ml EtOAc to remove unreacted chloranil, there was obtained 104 mg (39%) chloride pentahydrate.17 4',5,7-trimethoxyflavylium of $UV_{(0.01\,\text{N}\,\text{HC1}\,\text{m}\,\text{MeOH})}$: λ_{max} 242 (4.07), 277 (4.33), 324 (3.74) and 473 nm (4.60). Reported: 7 242 (3.96), 277 (4.29), 325 (3.72) and 474 (4.59). A portion of the product was converted to the FeCl₄ salt and recrystallized from AcOH. m.p. 186-7° (lit. 18 187°).

3',4',5,7-Tetramethoxyflavylium chloride (Tetramethylluteolininidin chloride). A mixture of 200 mg 4, 160 mg chloranil, and 5 ml HOAc were reacted and chromatographed as above. After freeze-drying and washing with EtOAc, there was obtained 131 mg (50%) of 3',4',5,7-tetramethoxyflavylium chloride as a red-orange solid. UV (001) HCI m MeOH): λ_{max} 241 (4.19), 279 (4.28), 3.20 (3.61) and 488 mm (4.57). Lit. λ_{max} 488 (4.62). A portion of the product converted to the FeCl₄ salt and recrystallized from AcOH had m.p. 206-7.5° (lit. λ_{max} 205-6°).

Acknowledgements—The authors are indebted to Dr. T. Radford, Mrs. M. Wilkinson, Mr. F. Shephard and Mr. L. Pope of our

laboratories for the determinations of MS, NMR and IR spectra.

REFERENCES

¹For a review of the early literature on anthocyanidin synthesis, see K. W. Bentley, *The Natural Pigments*, Interscience, New York (1960). The latest review of the field is by C. F. Timberlake and P. Bridle, *The Anthocyanins*, *The Flavonoids* (Edited by J. T. Mabry and H. Mabry) Part I, Academic Press, New York (1975).

²L. Jurd, Chem. & Ind. 1683 (1966).

³J. W. Clark-Lewis and R. W. Jemison, Aust. J. Chem. 23, 315 (1970).

⁴E. Haslam, The Chemistry and Biochemistry of Plant Proanthocyanidins, In Topics in Flavonoid Chemistry and Biochemistry (Edited by T. Farkas, M. Gabor and F. Kalley), Akademiai Kiado, Budapest (1975).

⁵R. E. Wrolstad and B. J. Struthers, *J. Chromat.* **55**, 405 (1971). ⁶L. Jurd, *J. Org. Chem.* **28**, 987 (1963).

⁷A. C. Waiss, Jr. and L. Jurd, Chem. & Ind. 743 (1968).

⁸D. D. Pratt and R. Robinson, J. Chem. Soc. 127, 166-75 (1925).
 ⁹H. L. Hergert, P. Coad and A. V. Logan, J. Org. Chem. 21, 304 (1956).

¹⁰K. Weinges, Ann. 615, 203 (1958).

¹¹J. W. Clark-Lewis and W. Korytnyk, J. Chem. Soc. 2367 (1958). ¹²A. K. Ganguly and T. R. Seshadri, Tetrahedron 6, 21 (1959).

¹³M. S. Kamat, P. Y. Mahajan and A. B. Kulkarni, *Ind. J. Chem.* 8, 119 (1970).

¹⁴B. R. Brown and M. R. Shaw, J. Chem. Soc. Perkin I, 2036 (1974).

¹⁵S. E. Drewes and D. G. Roux, *Biochem. J.* 92, 559 (1964).

¹⁶J. W. Clark-Lewis, Aust. J. Chem. 21, 2059 (1968).

¹⁷J. W. Gramshaw, A. W. Johnson and T. J. King, J. Chem. Soc. 4040 (1958).

D. D. Pratt, R. Robinson and P. N. Williams, *Ibid.* 125, 199 (1924).
 J. W. Clark-Lewis and R. W. Jemison, *Aust. J. Chem.* 21, 2247 (1968).